## CST Quick Guide rev20200401

1. Turn on the cytometer and log into your FACSDiVa account as usual.
2. All supplies are in white mini-fridge under FACSCalibur workstation.
2.1. Bead dilutions are stable $\geq 1$ week if kept refrigerated and protected from light.
2.2. Use existing tube of CST beads if volume is $\geq 300 \mathrm{ul}$ and age is $\leq 7$ days.
3. To make new tube of CST beads (all supplies in white mini fridge):
3.1. Use new 5 ml polystyrene FACS tube with cap
3.2. Label tube with "CST", lot\#, date: e.g. "CST 80296 3/30/20"
3.3. Vortex blue CST dropper bottle for 2-3s
3.4. Squeeze 1 drop CST bead stock into labeled tube
3.5. Add 600 ul DI H2O and vortex 2-3s
3.6. Immediately return blue CST dropper bottle to
4. In FACSDiVa, go to Cytometer Menu > CST.

5. A CST window will pop up, and it will take up to 1 min to connect. The bottom right corner will switch from "Connecting" to "Connected".


| O Connected |
| :--- |

6. In the new CST app window
6.1. Set Characterize to "Check Performance"
6.2. Lot ID matches blue dropper bottle " 80296 (RUO
6.3. Check the box for "Load Tube Manually".

7. Set fluidics to RUN, LOW, and rotate fine adjustment 5 full turns from either end.
8. Vortex bead tube and load on cytometer
9. Click the green "Run" button.

9.1. Select "yes" if bead lot has expired. They are stable for many years.

10. CST will detect beads and calculate results (can take up to 15 min .) If liquid in tube gets too low before the CST completes, click the red "Abort" button, make a new tube of beads as instructed in step 4 , and repeat steps 8-9.
11. When CST finishes, "Unload the CST bead tube" will pop up.
11.1. Select "OK".
11.2. Unload the tube and put it back to the fridge for the next user.
12. A generated PASS/FAIL report will show stats for each channel.
12.1. If fail, PRIME then rerun CST (using beads made today).
12.2. If fail again, email facs@ sbpdiscovery.org.

| Cytometer Performance Report |  |  |  |  |  |  |  |  |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| Cytometer: <br> Cytometer Name: <br> Serial Number: <br> Input Device: <br> Tube Loaded Manually: Cytometer Configuration: |  | LSRFortessa X20 <br> H656385091 <br> HTS <br> Yes <br> : SBPv2 2-Blue | 3-Red 6-Violet | 4-YGr3-UV | User: <br> Institution: <br> Software: <br> Date: <br> Cytometer P/F: | $\begin{array}{ll}  & \\ & \text { zsang } \\ & \text { Flow } \\ & \text { BD F/ } \\ & \text { se } \\ & \text { ne: } \\ \hline \end{array}$ | $17$ <br> ytometry CSDiva 8.0.1 2020 09:31 $20=39: 58$ | M (Expired) |
| Setup Beads |  |  |  |  |  |  |  |  |
| Bead Product: <br> Lot ID: <br> Bead Lot Information: |  | CST Setup Beads 80296 <br> Available | Part \#: 910858 <br> Expiration Date: $02 / 29 / 2$ |  | /2020 (Expired) |  |  |  |
| Detector Settings |  |  |  |  |  |  |  |  |
| Laser | Detector | Parameter | Target Value | Actual Target Value |  | \% Difference Target Value | Bright Bead \%Robust CV | Mid Bead Median Channel | Mid Bead \% Robust CV |
| Blue | FSC | FSC | 125000 | 125481 | 0 | 0.92 | 125556 | 0.92 |
| Blue | C | SSC | 125000 | 123950 | -1 | 3.06 | 124769 | 3.20 |
| Blue | B | B530 | 7532 | 7354 | -3 | 2.01 | 160 | 11.36 |
| Blue | A | R710 | 22887 | 22518 | -) | 264 | 617 | 1293 |

13. Close the window
14. 3 min bleach +3 min DI water clean, then proceed with your experiment.
